
Short Communication**Studies on moisture sorption isotherm and nutritional properties of dried Roselle calyces**

Ashaye, O. A.

*Obafemi Awolowo University, Institute of Agricultural Research and Training P.M.B
5029 Moor-Plantation Ibadan , Nigeria***Article history***Received: 22 May 2012**Received in revised form:**5 June 2012**Accepted: 20 June 2012***Abstract**

Freshly harvested roselle calyces of dark and light red varieties were sundried and oven dried and evaluated for proximate composition and moisture sorption studies. pH of sundried roselle calyces were higher than oven dried roselle calyces. Also, oven dried dark red Roselle calyces was significantly higher in crude protein, fat, dry matter, moisture and ash. Higher values potassium, phosphorus, sodium, magnesium, iron, zinc and calcium were also found in red Roselle calyces. Vitamin C content of oven dried light red Roselle calyces was significantly lower than Vitamin C of other Roselle calyces at $p < 0.05$. Roselle calyces exhibited the typical three stage sigmoidal curve found in most foods Braunaeur-Emett-Teller (BET Type II). There was also a concomitant increase in the equilibrium moisture content (EMC) as relative humidity increased in all roselle calyces irrespective of the temperature regimes. Sun dried roselle calyces gave the highest rate of water absorption unlike oven dried roselle calyces. In conclusion oven drying is the best method to dry freshly harvested roselle calyces, oven dried dark red roselle calyces was high in nutrient composition and all roselle calyces exhibited the typical three stage sigmoidal curve found in most foods Braunaeur-Emett-Teller (BET Type II).

Keywords*Roselle calyces**oven drying**sun drying**moisture sorption isotherm*

© All Rights Reserved

Introduction

Roselle (*Hibiscus sabdariffa* L.) is a member of the family *Malvaceae* to which okra, cotton and kenaf belong. The flowers of Roselle are generally small. Both the leaves and the fleshy base of the flower (the calyx) are employed in the preparation of soups and sauces. The vegetable is a popular diet during the raining season. Roselle calyx is a cheap source of vegetable protein, fat and minerals. Regular consumption of roselle may reduce nutritional deficiency problems such as night blindness, scurvy and rickets (Babalola *et al.*, 2001; Ashaye and Adeleke, 2009). Hygroscopy of foods affect storage, handling and processing. Valuable information of hygroscopy of foods can be obtained from moisture sorption characteristics of the food. Moisture sorption isotherm describes the equilibrium moisture content (E.M.C), which is the limit of the moisture that can be attained when food is exposed to air at a given temperature and water activity (Ashaye and Aina, 2008, Alakali *et al.*, 2010).

Information on equilibrium moisture content and

nutritional properties of dried roselle calyces will stand as a useful guide on its shelf life properties especially for food processors at culinary and industrial levels. This work evaluated the moisture sorption isotherm and nutritional properties of dried Roselle calyces

Materials and Methods*Raw materials*

Roselle calyces (*Hibiscus sabdariffa*) used for this research study were obtained from the experimental farm of Institute of Agricultural Research and Training, I.A.R. & T., Ibadan.

Preparation of samples

Fresh samples of red and light red roselle calyces were sun dried for three days, oven dried for 24hrs at 50°C and then packed for analysis.

Determination of moisture content

One (1) gram of each sample was weighed using mettle pc 4410 balance into dry pre-weighed crucibles. The samples were dried in the oven at 50°C

*Corresponding author.

Email: kayodeashaye@yahoo.com

overnight and were cooled. The percentage of dry matter was calculated by using formula below:

$$\text{Dry matter \%} = \frac{\text{Dry matter weight}}{\text{Weight of sample before oven}} \times 100$$

Also, the percentage of moisture content was determined by using the formula below:

$$\% \text{ Moisture content} = 100 - \% \text{ Dry matter}$$

Determination of ascorbic acid

Ascorbic acid was determined using the procedure described by Kirk and Sawyer (1991). Standard indophenol's solution was prepared by dissolving 0.05 g 2,6-dichloro Indophenol in water diluted to 100 ml and filtered. To standardize, 0.053 g of ascorbic acid was dissolved in 90 ml of 20% metaphosphoric acid and diluted with water to 100 ml. 10 ml of this solution was pipette into a small conical flask and titrated with indophenol's solution until a faint pink colour persists for 15 seconds. 2 ml of the extracted juice from the calyces was pipette into a conical flask and 5 ml of 20% metaphosphoric acid (as stabilizing agent) was added and made up to 10 ml mark with water. It was titrated with the indophenols solution a faint pink colour persists for 15seconds. The vitamin content in the calyces was calculated

$$\text{Vitamin C in mg/100g} = \frac{\text{Titre value} \times 0.212 \times 100}{\text{Wt of sample}}$$

pH determination

The pH meter (model BA 350 EDT instruments) was standardized with standard buffer solution 4. 0.and 7.0. The pH was measured by inserting directly the electrodes into 10ml beaker containing the sample.

Determination of ash

The sample (2 g) was weighed into a porcelain crucible. This was transferred into the muffle furnace set at 550°C and left for about 4 hours. About this time it had turned to white ash. The crucible and its content were cooled to about 100°C in air, then room temperature in a desiccator and weighed (AOAC, 1990). The percentage ash was calculated from the formula below:

$$\% \text{ Ash content} = \frac{\text{Weight of ash}}{\text{Original weight of sample}} \times 100$$

Determination of crude protein

The micro-Kjeldahl method for protein determination was employed for protein determination. The finely ground dried sample (0.5 g) was weighed into the micro-Kjeldahl flask. To this were added 1

Kjeldahl catalyst tablet and 10 ml of conc. H₂SO₄. These were set in the appropriate hole of the digestion block heaters in a fume cupboard. The digestion was left on for 4 hours after which a clear colourless solution was left in the tube. The digest was carefully transferred into 100 ml volumetric flask, thoroughly rinsing the digestion tube with distilled water and the volume of the flask made up to the mark with distilled water. 5 ml portion of the digest was then pipetted to Kjeldahl apparatus and 5 ml of 40% (w/v) NaOH added.

The mixture was then steam distilled and the liberated ammonia collected into a 50 ml conical flask containing 10 ml of 2% boric acid plus mixed indicator solution. The green colour solution was then titrated against 0.01 NHCl solution. At the end point, the green colour turns to wine colour, which indicates that, all the nitrogen trapped as ammonium borate have been removed as ammonium chloride. The percentage nitrogen was calculated by using the formula:

$$\% \text{ N} = \frac{\text{Titre value} \times \text{atomic mass of nitrogen} \times \text{normality of HCl used} \times 4}{\text{Volume of HCl used}}$$

The crude protein is determined by multiplying percentage nitrogen by a constant factor of 6.25 (AOAC, 1990).

Crude fat determination

The dried sample (1 g) was weighed into fat free extraction thimble and plug lightly with cotton wool. The thimble was placed in the extractor and fitted up with reflux condenser and a 250 ml soxhlet flask which has been previously dried in the oven, cooled in the dessicator and weighed. The soxhlet flask is then filled to ¾ of it volume with petroleum ether (b.pt. 40 – 60°C) and the soxhlet flask extractor plus condenser set was placed on the heater. The heater was put on for six hours with constant running water from the tap for condensation of ether vapour. The set is constantly watched for ether leaks and the heat sources is adjusted appropriately for the ether to boil gently. The ether is left to siphon over several times at least 10 – 12 times until it is short of siphoning. It is after this is noticed that any ether content of the extractor is carefully drained into the ether stock bottle. The thimble-containing sample is then removed and dried on a clock glass on the bench top. The extractor flask with condenser is replaced and the distillation continues until the flask is practically dried. The flask which now contains the fat or oil is detached, its exterior cleaned and dried to a constant weight in the oven (AOAC, 1990). If the initial weight of dry soxhlet flask is W_o and the final weight

of oven dried flask + oil/fat is W_1 , percentage fat/oil is obtained by the formula:

$$\frac{W_1 - W_n}{\text{Weight of sample taken}} \times \frac{100}{1}$$

Crude fibre determination

The sample (2 g) was accurately weighed into the fibre flask and 100 ml of $0.25\text{NH}_2\text{SO}_4$ added. The mixture was heated under reflux for 1 hour with the heating mantle. The hot mixture was filtered through a fibre sieve cloth. The filtrate obtained was thrown off and the residue was returned to the fibre flask to which 100 ml of (0.31NNaOH) was added and heated under reflex for another 1 hour.

The mixture was filtered through a fibre sieve cloth and 10ml of acetone added to dissolve any organic constituent. The residue was washed with about 50 ml hot water twice on the sieve cloth before it was finally transferred into the crucible. The crucible and the residue was oven-dried at 105°C overnight to drive off moisture. The oven-dried crucible containing the residue was cooled in a dessicator and later weighed to obtain the weight W_1 . The crucible with weight W_1 was transferred to the muffle furnace for ashing at 550°C for 4 hours. The crucible containing white or grey ash (free of carbonaceous material) was cooled in the dessicator and weighed to obtain W_2 . The difference $W_1 - W_2$ gives the weight of fibre (AOAC, 1990). The percentage fibre was obtained by the formula:

$$\% \text{ Fibre} = \frac{W_1 - W_2}{\text{Weight of sample}} \times \frac{100}{1}$$

Phosphorus determination

The ash of each sample obtained was treated with 2M HCl solution 10 ml of the filtrate solution was pipetted into 50 ml standard flask and 10 ml of vanadate-molybdate yellow solution was added and the flask was made up to mark with distilled water, stoppered and left for 10 min for full yellow colour development. The concentration of phosphorus was obtained by measuring the optical density (OD) or absorbance of the solution in a Spectronic 20 spectrophotometer or calorimeter at a wave length of 470mm. The percentage phosphorus was calculated from a standard graph by appropriate mathematical relationship applicable to such determination (AOAC, 1990).

Potassium and sodium

Potassium and Sodium were estimated using a Jenway digital flame photometer spectronic 20.. (AOAC,1990)

Calcium, magnesium, zinc and Iron

Calcium, magnesium, zinc and iron were determined spectrophotometrically using Bulk 200 atomic absorption spectrophotometer (Buck Scientific, Norwalk) and the absorption compared with absorption of standards of these minerals (Osundahunsi and Aworh, 2003).

Determination of equilibrium moisture content (EMC)

Samples were conditioned to constant weights over either 90% concentrated sulphuric acid (drying) before the EMC was determined. A static gravimetric method (Spiess and Wolf, 1983) was used. Duplicate sample, 1 g each were placed in the upper section of each glass desiccators on wire mesh, while the lower section contained standard salt solution over excess salt. After inserting the samples and salts, the desiccators were seated with silicone grease and placed in a constant temperature environment of 18°C . The samples were weighed at interval of 24 hours until equilibrium was reached when four consecutive measurements are the same. This took between 20 – 25 days. The EMC was determined by Labuza (1984) and average values were used for all calculations. The salt used and their corresponding water activity (a_w). EMC values were obtained using equation below:

$$\text{EMC} = \frac{\text{WF} - \text{Wi} (\text{Wt of sample} \times \text{moisture content})}{\text{Sample Wt} - \text{MC}}$$

Wf = Final weight of sample + crucible

Wi = Initial weight of sample + crucible.

Statistical analysis

Data was subjected to analysis of variance and their means were separated by Duncan Multiple range test Duncan (1955).

Results

Proximate composition of oven and sun dried roselle calyces

In Table 1, it can be depicted that the pH of sundried roselle calyces were higher than oven dried roselle calyces. Also, oven dried dark red Roselle calyces was significantly higher in crude protein, fat, dry matter, moisture and ash Higher values potassium, phosphorus, sodium, magnesium, iron, zinc and calcium were also found in red Roselle calyces. Vitamin C content of oven dried light red Roselle calyces was significantly lower than Vitamin C of other Roselle calyces at $p < 0.05$

Table 1. Chemical composition of oven and sun-dried roselle calyces

	pH	Crude Protein (%)	Fat (%)	Dry Matter (%)	Moisture (%)	Crude Fibre (%)	Ash (%)	K (%)	P (%)	Na (%)	Mg (%)	Fe (%)	Zn (%)	Ca (%)	Vit. C Mg/100g
Sun dried Dark Red Roselle	2.56d	1.575b	0.57b	84.42c	15.58b	2.48b	8.79b	0.078c	0.059b	0.0113c	0.029c	0.0038c	0.0014c	0.168c	14.84a
Sun dried Light Red Roselle	2.68c	0.875d	0.19d	83.26d	16.74a	1.61d	8.14d	0.072d	0.017d	0.0108d	0.021d	0.0034d	0.0009d	0.155d	11.66a
Oven dried Dark Red Roselle	3.48b	2.10a	0.61a	93.75a	6.25d	2.69a	8.97a	0.092a	0.072a	0.0134a	0.043a	0.0057a	0.0021a	0.23a	11.66b
Oven dried Light Red Roselle	3.62a	1.225c	0.23c	92.69b	7.31c	1.74c	8.32c	0.083b	0.026c	0.0119b	0.031b	0.0045b	0.0016b	0.18b	8.48c

Means in the same column followed by the same letter are not significantly different from each other at $P < 0.05$

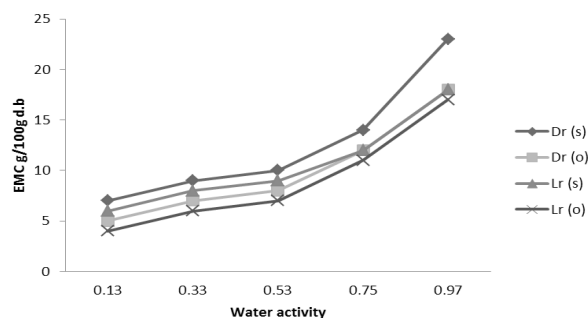


Figure 1. Water adsorption property of dried roselle calyces at 270C. Dr (s): Sun dried dark red roselle calyce, Dr (o): Oven dried dark red roselle calyce. Lr (s): Sun dried light red roselle calyce. Lr (o): Oven dried light red roselle calyce

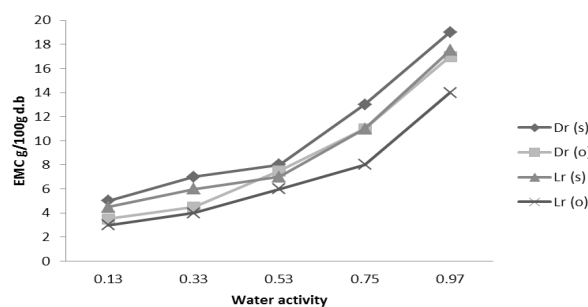


Figure 2. Water adsorption property of dried roselle calyces at 350C. Dr (s): Sun dried dark red roselle calyce, Dr (o): Oven dried dark red roselle calyce, Lr (s): Sun dried light red roselle calyce, Lr (o): Oven dried light red roselle calyce

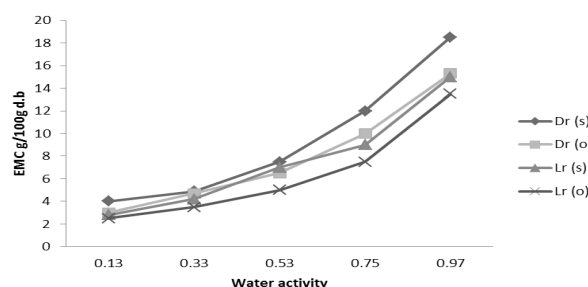


Figure 3. Water adsorption property of dried roselle calyces at 370C. Dr (s): Sun dried dark red roselle calyce, Dr (o): Oven dried dark red roselle calyce, Lr (s): Sun dried light red roselle calyce, Lr (o): Oven dried light red roselle calyce

Water adsorption isotherm of sun dried and oven dried roselle calyces

Figures (1, 2 and 3) show the rate of water adsorption of sun dried and oven dried roselle calyces at 27°C, 35°C, and 37°C. The roselle calyces exhibited the typical three stage sigmoidal curve

found in most foods Braunaeur-Emett-Teller (BET Type II). There was also a concomitant increase in the equilibrium moisture content (EMC) as relative humidity increased in all roselle calyces irrespective of the temperature regimes. Sun dried roselle calyces gave the highest rate of water absorption unlike oven dried roselle calyces.

Discussion

This study evaluated the moisture sorption isotherm and nutritional properties of dried roselle calyces. The significance differences observed in the nutrient composition of the dried roselle calyces may be due to the increased activities of the microorganisms due to chance fermentation resulting in the extracellular enzymatic secretion and production of organic acids from available nutrients as described by (Okafor, 1978; Ashaye *et al.*, 2008; Bolade *et al.*, 2009). Also varietal characteristics, soil nutrient and climatic condition may be responsible for these observation (Ho-Hsien *et al.*, 2005). Differences in Vitamin C content of the calyces is due to instability of Vitamin C at higher temperatures (Ashaye *et al.*, 2008)

It can be depicted from the moisture sorption studies that it is not advisable to store these roselle calyces at relative humidity's greater than 60% because of greater rate of water uptake. Maintaining relative humidity between 60% and 62% will be safe for storage purposes. This trend agrees with published studies of (Ajibola *et al.*, 2003; Denloye and Adejohn, 1983).

Conclusion

It can be concluded from this work that oven drying is the best method to dry freshly harvested roselle calyces, oven dried dark red roselle calyces had better nutrient quality and all roselle calyces exhibited the typical three stage sigmoidal curve found in most foods Braunaeur-Emett-Teller (BET Type II) with relative humidity range of 60-62% as safe relative humidity for storage irrespective of elevated temperatures.

References

- Alkali, J.S., Irtwange, S.V. and Agbali, F.A. 2010. Physico thermodynamic properties of soy-gari. *Journal of Food Processing and Preservation* 34: 470-489.
- AOAC, 1990. Association of Official Analytical Chemists. *Official Methods of Analysis*. 15th Edition Washington D.C.
- Ajibola, O.O. Aviara, N.A. and Ajetumobi, O.E. 2003. Sorption equilibrium and thermodynamic properties of cowpea (*Vigna unguiculata*). *Journal of Food Engineering* 54: 317-324.
- Ashaye, O.A. and Aina, J.O. 2008. Moisture sorption characteristics of cowpeas. *African Journal of Pure and Applied Sciences* 1: 39-41.
- Ashaye, O.A. Raji, J.A. and Afolabi, O.O. 2008. Roselle tea production and storage. *African Journal of Pure and Applied Sciences*.1: 109-115.
- Ashaye, O. A. Adeleke, T.O. 2009. Quality attributes of stored Roselle Jam. *International Food Research Journal* 16: 363-371.
- Babalola, S.O. Babalola, A.O. and Aworh, C.O. 2001. Compositional attribute of the calyces of Roselle (*Hibiscus sabdariffa* var *sabdariffa* L). *The Journal of Food Technology in Africa* 6: 133-134.
- Bolade, M.K. Oluwalana, I.B. and Ojo, O. 2009. Commercial practice of roselle (*Hibiscus sabdariffa* L.) beverage production: Optimization of hot water extraction and sweetness level. *World Journal of Agricultural Sciences* 5(1): 126-131.
- Denloye, A.O. and Adejohn, A.O. 1983. Moisture sorption isotherms of some Nigerian food grains. *Journal of Stored Product Research* 21: 53-58.
- Duncan, D.B. 1955. Multiple Range and Multiple F Tests *Biometrics*, 11: 1-5.
- Ho-Hsien, C. Pi-Jen, T. Shyn-Huang, C. Yu-Ming, S. Chao-Chin, C. and Tzou-Chi, H. 2005. Grey relational analysis of dried roselle (*Hibiscus sabdariffa* L.). *Journal of Food processing and preservation* 29: 228-245.
- Kirk, R.S. and Sawyer, R. 1991. *Pearson's composition and analysis of food*, 9th Ed. Longman, Singapore 221-225.
- Labuza, T.P. 1984. *Moisture sorption: practical aspects of isotherm measurement and use*. St. Paul, Minnisota. American Association of Cereal Chemists. 2-8
- Okafor, N. 1978. . *Microbiology and Biochemistry of oil palmwine*. *Advances in Applied Microbiology* 24: 237-256
- Osundahunsi, O.F. and Aworh, O.C. 2003. Nutritional evaluation with emphasis on protein quality of maize-based complementary foods enriched with soybean and cowpea tempe. *International Journal of Food Science Technology* 38: 809-813.
- Spiess, W.E.L. and Wolf, W. 1983. The results of the COST 90 project on water activity. In: *Physical properties of foods*, pp. 65-87. Jowitt, R., Escher, F., Hallstrom, B., Meffert, H.F.Th., Spiess, W.E.L., and Vos, G. (Eds.), Elsevier Science Publishing Company, London, England.