Short-term consumption of Gelam honey reduces triglyceride level

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Abstract

The study was carried out to evaluate short-term administration of Gelam honey. A single oral administration of the honey at a dose of 5000 mg/kg body weight on male Sprague Dawley rats (test group) for 14 days did not produce any signs of toxicity, behavioral changes, mortality, changes on gross appearance or histopathological changes of internal organs. The examinations of signs, animal behavior and health monitoring showed no abnormalities in the test group as compared to the rats unfed with the honey (control group). The test group had progressive increased both body weight and in the meal pattern analysis. However, triglycerides level was found significantly decreased in the test group. It suggested that the honey might have a decent effect in controlling the blood triglyceride level. Polyphenol contents in the honey may play the role to reduce the triglyceride level. Biochemical test for aspartate aminotransferase (AST), alanine transaminase (ALT), urea, creatinine, cholesterol and glucose of rats in the test group were in the normal range compared to the control. There were no significant changes in the absolute and relative organ weight between the two groups. As a conclusion, tested dose of Gelam honey is safe and has medical potential. Meanwhile, lethal dose (LD$_{50}$) of the honey was found to be greater than 5000 mg/kg body weight. Long period of Gelam honey consumption should be conducted to observe and confirm those effects.

Introduction

Honey is sweet in taste, golden in color and concentrated liquid in natural condition. It is obtained from liquid nectar of many plants and is a food source for honey bees (Bogdanov et al., 2012). Honey is classified from clear to dark color due to its flavors, mineral contents, quality and floral source (Uthurry et al., 2011). Physical characteristic of honey is according to its viscous liquids and able to be identified by its flavor, appearance, color, crystallization and presence of pollen grains in honey sediment (Bogdanov et al., 2012).

Gelam honey is one of many types of monofloral Malaysian honey, which is produced by honey bee from Melaleuca species plant (Hussein et al., 2011). It has high components of polyphenols and non-phenol content activities compared to other local honeys in Malaysia (Piljac-Zegarac, 2009). Thus, it possesses both antioxidant and polyphenols agents, which are important to prevent occurrence of chronic diseases, infections and can improve enzyme activities to the optimal level in the body (Alvarez-Suarez et al., 2010).

Honey is claimed does not give any adverse effect in daily intake; because honey is a natural product that has biological and chemical bioequivalent toward human body needs. However, only few studies have to date taken into cognizance of the possible toxicity effects of Gelam honey even though progressive researches have been conducted on the honey. In addition, toxicity study is one of the requirements towards to clinical studies in order to observe other possible medicinal value of honey systematically. Therefore, this study was undertaken to observe the toxicity effects of acute administrations of Gelam honey at the highest doses 5000 mg/kg body weight on male Sprague Dawley rats for 14 days according to the Organisation for Economic Co-operation and Development (OECD).

Materials and Methods

Sample collection

Gelam honey was collected from Gelam forest, Terengganu, Malaysia. The sample was irradiated...
with 25 kGy gamma irradiation using radioactive sources cobalt 60 (model JS8900) at Malaysian Nuclear Agency (MINT), Selangor, Malaysia (Hussein et al., 2011). The irradiated honey was then kept in 4°C, away from direct sunlight in amber bottles.

**Experimental animals**

The experimental protocol was approved by the Research Committee on The Ethical Use of Animal (UiTM Care) Reference No. 05/2012. Ten apparently healthy male Sprague Dawley rats were obtained from Laboratory Facilities of Animal Management (LAFAM), University Teknology MARA (UiTM), Puncak Alam, Selangor, Malaysia. At the commencement of its dose, each animal was eight weeks old and its weight was between 180 to 220 g (OECD/OCDE, 2001). The rats were housed at one rat per cage and maintained in standard environmental conditions under an ambient temperature of 25 ± 2°C and 40 - 65% relative humidity, with a 12-h light/dark cycle. They were fed with certified rodent food (Rodent Diet Specialty Feeds, Glen Forrest Australia) and drinking water was available ad libitum. The animals were acclimatized for five days prior to the commencement of the study and labeled appropriately. A completely randomized design was used in dividing the rats into two groups that consisted five male in each group i.e. control rats were orally administered with distilled water and rat chow ad libitum (control group) and tested group where rats were fed with Gelam honey (5000 mg/kg body weight) (test group).

**Acute toxicity study**

Acute oral toxicity test was performed to observe short-term administration of honey according to the guidelines of OECD for testing of chemicals, TG 423 with slightly modifications (OECD/OCDE, 2001). It measures adverse effects occurred following oral administration of a single dose of a substance, or multiple doses given within 24 hours such as general behavior, respiratory pattern, cardiovascular signs, motor activities, reflexes and changes in skin and fur texture. For the test group, fixed dose of Gelam honey, 5000 mg/kg body weight of male Sprague Dawley rats were orally given once whereas the control group was administered with distilled water. The doses given were calculated according to animal’s body weight on the week of specified treatment. The median lethal oral dose (LD_{50}), which is a statistically derived single dose of a substance that can be expected to cause death in 50% of animals when administered by the oral route, was determined. The LD_{50} value is expressed in terms of weight of test substance per unit weight of test animal (mg/kg)(OECD/OCDE, 2001). Dosing for this study was initiated at 5000 mg/kg because there was no estimate of the substance’s lethality available so far. Thus, the maximal dose was used as suggested by the OECD. Meanwhile, mortality and the clinical signs of toxicity were observed at 30 minutes, 1, 2 and 4 hours and thereafter once a day for the next 14 days (Samat et al., 2014).

**Body weight and meal pattern analysis**

The body weight of each rat was recorded daily and the differences of the body weight were noted (OECD/OCDE, 2001). The food efficiency was calculated at the end of study as the quantity of food (in g) consumed by each rat by subtracting the weight of uneaten food from initial weight of food (Vinicus et al., 2006). Then, the total number of kilocalories that each rats consumed was determined by multiplying the caloric content of 1 g of each diet by the total quantity eaten (Mohd Saleh et al., 2012).

**Measurement of blood chemical parameters**

The nonheparinized blood was subjected to biochemical tests using an Auto Analyser ILAB 300 Plus Clinical Chemistry Analyser, Milano Italy. The blood was allowed to coagulate prior to centrifuged. Then, the serum was separated to analyze glucose, urea, creatinine, aspartate aminotransferase (AST), alanine transaminase (ALT), glucose, triglycerides and cholesterol.

**Histology evaluation**

On day 14 of the dosing period, all the animals were euthanized by diethyl ether. The organs such as heart, liver, spleen, kidneys and lungs were weighed and relative organ weights (ROW) were calculated (Mohd Saleh et al., 2012). The ROW of each animal was then calculated as follows:

\[
\text{Relative organ weight} = \frac{\text{Absolute organ weight (g) / Body weight of rat on sacrifice day (g)}}{100}
\]

For histology evaluation, the internal organs were fixed in 10% neutral buffered formalin, trimmed and a 5 μ thickness of tissue sections were stained with hematoxylin and eosin for histopathological investigation.

**Statistical analysis**

Results were expressed as mean ± standard average mean (SEM). Statistical significance was determined by one-way analysis of variance.
During 14 days of diet regimens, cumulative food intake of rats among both groups was slightly different in mass. Mean total food intake for the test and control groups were 358.18 g and 325.25 g, respectively. In sum, total calories consumed from the mean total food intake for rats fed with Gelam honey was 2763.79 calories compared to the control group was 2509.62 calories (Figure 1(c)).

**Biochemical analysis**

Overall, results of biochemical analysis for the test group showed an increased but still in normal range for all tests (ALT, urea, creatinine, cholesterol and glucose) except for AST showed significantly increased compared to the control (Table 1). However, for triglyceride level, rats fed with Gelam honey showed the significantly decreased (0.92±0.14 mmol/L) compared to the control (1.33±0.12 mmol/L).

**Relative organ weight**

Relative organ weight for the liver, kidney, spleen, heart and lung were not affected by Gelam honey supplementation (Table 2). In addition, gross necropsy findings did not reveal changes in any of the organs.
Histology evaluation

For the histology investigation, no pathological changes were observed in the liver of animals in the control group (Figure 2(a)) and the test group (Figure 2(b)), which showed normal lobular architecture with central vein and radiating hepatic cords. The macroscopic observation of the organs did also not present any significant morphological or hemorrhagic changes due to the administration of Gelam honey at 5000 mg/kg body weight. Other organs including spleen, lung, kidney and heart showed no sign of pathological changes compared with the corresponding organs of the control.

Table 1. Biochemical changes of male Sprague Dawley rats after 14 days of acute oral administration of Gelam honey (5000 mg/kg). Results are reported as mean ± SEM (n=5); ALT: alanine aminotransferase; AST: aspartate aminotransferase. * significant different (p < 0.05) versus control.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control group</th>
<th>Test group</th>
<th>Reference values</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>52.90 ± 3.23</td>
<td>55.26 ± 3.52</td>
<td>52.70 ± 4.20</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>103.56 ± 2.36</td>
<td>123.96 ± 2.34</td>
<td>108.20 ± 12.10</td>
</tr>
<tr>
<td>AST (U/L)</td>
<td>38.60 ± 1.73</td>
<td>40.50 ± 0.73</td>
<td>41.10 ± 9.11</td>
</tr>
<tr>
<td>Urea (mg/dL)</td>
<td>70.82 ± 2.56</td>
<td>68.98 ± 1.98</td>
<td>70.90 ± 8.20</td>
</tr>
<tr>
<td>Creatinine (μmol/L)</td>
<td>1.49 ± 0.19</td>
<td>1.27 ± 0.13</td>
<td>1.52 ± 0.19</td>
</tr>
<tr>
<td>Cholesterol (mmol/L)</td>
<td>1.33 ± 0.12</td>
<td>0.92 ± 0.14</td>
<td>1.33 ± 0.51</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>4.26 ± 0.32</td>
<td>4.40 ± 0.19</td>
<td>4.30 ± 0.39</td>
</tr>
</tbody>
</table>

(ANOVA) followed with Tukey’s test post-hoc using SPSS software version 18.0 (SPSS, Chicago IL, USA). Values with a confidence level of p ≤ 0.05 were considered as significant.

Table 2. Effects of Gelam honey and control on relative organ weights (ROW) of male Sprague Dawley rats. Results are reported as mean ± SEM (n=5); * significant different (p < 0.05) versus control.

<table>
<thead>
<tr>
<th>Organ</th>
<th>Control group</th>
<th>Test group (Gelam honey)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Heart</td>
<td>0.202 ± 0.03</td>
<td>0.208 ± 0.03</td>
</tr>
<tr>
<td>Spleen</td>
<td>0.149 ± 0.01</td>
<td>0.178 ± 0.03*</td>
</tr>
<tr>
<td>Lung</td>
<td>0.288 ± 0.03</td>
<td>0.275 ± 0.04</td>
</tr>
<tr>
<td>Kidney</td>
<td>0.530 ± 0.02</td>
<td>0.522 ± 0.04</td>
</tr>
<tr>
<td>Liver</td>
<td>2.930 ± 0.21</td>
<td>2.810 ± 0.25</td>
</tr>
</tbody>
</table>

Discussion

Honey has been used for medicinal purposes in many cultures since the ancient times (Enette Larson-Meyer et al., 2010; Ajibola et al., 2012). It is one of the oldest and most enduring-substance used in wound management. Scientific evidences for their efficacy are widely studied but systemic safety studies are still lacking. Therefore, it is essential to evaluate the toxicity of honey in animal model to ensure its safety and may further it in clinical studies.

The study was designed to assess the acute toxicity induced by Gelam honey after administrated orally at the maximum dose 5000 mg/kg body weight of Sprague Dawley rats based on OECD guidelines. According to the OECD 423 test guidelines (Jonsson...
et al., 2012), the 5000 mg/kg body weight doses were chosen because the dosage is safe, applicable and appropriate to evaluate safety of some substances.

The acute oral toxicity study is performed to observe a short period consumption of substances. The present study was conducted by administering the single dose of Gelam honey for 14 days using male Sprague Dawley rats only. Male rat is an appropriate model for the short course study since female rat is frequently affected by its hormonal levels (Mark et al., 2005). There were no any detectable physical toxicological symptoms like mortality, morbidity, loss of fur, diarrhea, convulsion, salivation, fatigue, sleep, coma or aggression. The aforementioned signs are indicators of interference of toxicants with the passage of electrical impulses down the axon.

The increment of body weight for the test group considered as normal when compared to the control group. Then, the increasing patterns of food intake for the test group paralleled with calories consumed. Although, biochemical test for hepatic function (ALT and AST) were slightly higher, gross pathological examination of the Gelam honey groups (Figure 4(b)) did not reveal any abnormalities, presence of lesions or changes in the color of the internal organs compared to the control (Figure 2(a)).

Intriguingly, triglycerides levels in rats fed with the honey were significantly decreased compared to the control. It indicates Gelam honey consumption might not be having lipogenic effects and reveals its potential in controlling triglycerides level. The major compound contributes to the effects are polyphenols since the compounds were reported available in the honey (Alvarez-Suarez et al., 2010; Hussein et al., 2011). Polyphenols were widely reported to control triglyceride level, consequently provide benefits to the cardiovascular system (Khurana et al., 2013).

There were no significant differences in internal organ weight. Organ weight measurement is important to access general toxicity because any changes in organ weight is a sensitive indicator for toxicity (Norazmir and Ayub, 2010). ROW results of liver were slightly decreased compared to control but not significant (Mark et al., 2005; Mark et al., 2012). ROW of liver was measured, as it is the target organ of the most toxicants. Normally, toxicants enter the body via the gastrointestinal tract and are carried by the hepatic portal vein to the liver for elimination (Srivinas et al., 2010).

The findings imply that short-term exposure to high concentration of Gelam honey does not produce health hazards in the male Sprague Dawley rats. The slightly changes observed in the body weight and body weight gain may have resulted from physiological changes in rats such as metabolism, food and water intake. At the same time, Gelam honey consumption may have medical benefit in controlling triglyceride level in the bloodstream.

Conclusion

Gelam honey at the highest concentration level tested (5000 mg/kg body weight of male Sprague Dawley rats) did not cause any mortality, physiological or behavioral changes in rats. The results suggest that the Gelam honey does not demonstrate acute toxicity effect in rats even at the maximal concentration based on the OECD guidelines. It may have good effects in fatty acid metabolism via controlling triglycerides level in the bloodstream through its polyphenol compounds. Further studies to assess a long-term safety of the Gelam honey and its effect on the lipid metabolism are intensively carrying out in laboratory.
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References


