

Review

Health effects of omega-3 polyunsaturated fatty acids in common diseases¹Jia, G., ¹Qiong, Z. and ^{1,2*}Yong-Hua, W.¹Guangdong Yue-s Special Nutrition Technology Co., Ltd., Foshan, 528000, Guangdong, China²School of Food Sciences and Engineering, South China University of Technology, Guangzhou, 510641, Guangdong, China**Article history**

Received: 19 October 2020

Received in revised form:

9 March 2021

Accepted:

12 March 2021

Abstract

Omega-3 polyunsaturated fatty acids (n-3 PUFAs), such as alpha-linolenic, eicosapentaenoic, and docosahexaenoic acids mostly exist in marine-derived foods, and have shown beneficial effects for hypertriglyceridemia, endothelial function, inflammation, and oxidative stress. Studies suggest that n-3 PUFAs can regulate the activity of NF-κB, Nrf2, SREBP-1c, and PPARα, which are linked to inflammations, ROS homeostasis, and lipid metabolism. Several epidemiological trials and physiological studies indicated protective effect of n-3 PUFAs against various common diseases such as cardiovascular diseases, diabetes mellitus, and non-alcoholic fatty liver disease. This review summarises the findings of many such studies highlighting the beneficial effects of n-3 PUFAs.

Keywords

n-3 PUFAs,
beneficial effect,
disease risk factors,
common diseases

© All Rights Reserved

Introduction

Polyunsaturated fatty acids (PUFAs) carry several double bonds. As the name suggests, the first double-bond in omega-3 polyunsaturated fatty acids (n-3 PUFAs) is at the third carbon from the methyl end. Alpha-linolenic (ALA), eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA) are the three major n-3 PUFAs (Calder, 2012). ALA is mainly produced in plants such as flaxseed, canola, and butternut, while EPA and DHA are mostly produced in marine animals such as fish, algae, and seals. Humans cannot endogenously synthesise n-3 PUFAs, and therefore mainly depend on diet sources. In humans, ALA can be biochemically converted into EPA and DHA; however, the conversion rate is very poor (ALA to EPA: 0.2 - 8%; ALA to DHA: 0 - 4%) (Mozaffarian and Wu, 2011). Due to this, marine food has emerged as an important direct source of EPA and DHA. Additionally, the fish oil in the form of ethyl esters (EEs) or acylglycerols is a popular commercially available n-3 PUFA supplement. Likewise, other marine-sourced oils, such as algal, fungal, and krill oils have been commercially accepted for their rich n-3 PUFA content (Shahidi and Ambigaipalan, 2018).

Various *in vitro*, animal, and clinical studies have shown antioxidant, anti-inflammatory, and cardiovascular-regulatory functions of n-3 PUFAs (Mozaffarian and Wu, 2011). Accordingly, n-3 PUFAs are conceived as healthy fats. In 2007, the American Dietetic Association (ADA) and Dietitians

of Canada (DC) declared that the basal n-3 PUFA requirement of 500 mg/day in adults is equivalent to 8 oz. of fish per week (Kris-Etherton *et al.*, 2007). In 2013, the DC again emphasised at least twice per week consumption of fish diet approximately provides 0.3 - 0.45 g n-3 PUFAs each day (Shahidi and Ambigaipalan, 2018).

Given the numerous physiological functions of n-3 PUFAs, they have been extensively studied for their protective effect against various diseases (Innes and Calder, 2020). This review thus aims to briefly summarise the beneficial effects of n-3 PUFAs with potential underlying mechanisms in several diseases.

n-3 PUFAs alleviate potential disease risk

Dietary triglycerides (TGs) enter the blood circulation from the small intestine as chylomicrons, and get hydrolysed by lipoprotein lipase (LPL) into free fatty acids (FFA), which are then consumed as the energy source by various cells such as muscle cells, while the excess TG is stored in the liver. In case of increased energy demand, hepatic TG combined with apolipoprotein B-100, also known as very-low-density lipoprotein (VLDL), is secreted back into the blood, where LPL can transform it into LDL for muscle cells (Alves-Bezerra and Cohen, 2017). High TG level, mostly a result of unhealthy lifestyle and diet, can cause endothelial dysfunction, which leads to decreased flow-mediated dilatation of blood vessels which could progress to other disorders (Reiner, 2017). Therefore, a high TG level is

*Corresponding author.
Email: yonghw@scut.edu.cn

considered a serious health risk factor. In 2016, European data concluded that individuals with high TG level of 6.6 mmol/L (580 mg/dL), when compared with individuals with healthy TG level of 0.8 mmol/L (70 mg/dL), have multiple fold higher risk of myocardial infarction (5.1-fold), ischemic heart disease (3.2-fold), ischemic stroke (3.2-fold), and all-cause mortality (2.2-fold) (Nordestgaard, 2016). Moreover, the oxidised TG-LDL triggers the secretion of inflammatory cytokines by macrophages which initiate a series of innate and adaptive immune responses (Rhoads and Major, 2018). Notably, several high-TG-level-induced diseases also involve oxidative stress, which can further aggravate the disease progression (Peverill *et al.*, 2014). Many studies reported that the beneficial effect of n-3 PUFAs can contest these negative factors, including TG accumulation, inflammatory response, and oxidative stress. Next, we summarise these related findings.

Hyperlipidaemia

Hyperlipidaemia (TG level ≥ 1.7 mmol/L or ≥ 150 mg/dL) is often associated with secondary disorders such as cardiovascular disease (CD) and/or type 2 diabetes mellitus (T2DM). Approximately 25% of the US adult population is hyperlipidemic, while severe hyperlipidaemia (≥ 5.6 mmol/L or ≥ 500 mg/dL) is a well-established initiation factor for secondary diseases (Toth, 2016). Several studies showed that n-3 PUFAs can lower the TG level in hyperlipidemic individuals. Zeman *et al.* (2006) showed that hyperlipidemic individuals who received n-3 PUFAs for three months showed a significant decrease in the plasma TG level than the individuals of the control group. Similarly, Zhu *et al.* (2008) showed a remarkable decrease of plasma TG level among hyperlipidemic participants after 24-week administration of n-3 PUFAs. Chan *et al.* (2016) also showed similar results. Besides, a study of 176 hyperlipidemic subjects, which were randomly assigned into the placebo-controlled, 1, 2, and 3 g n-3 PUFAs treatment groups, showed a

dose-dependent decrease in plasma TG level in a 2-month follow-up study (Oh *et al.*, 2014).

n-3 PUFAs can lower the plasma TG level via three major pathways: (1) n-3 PUFA supplementation inhibits the activity of sterol receptor element-binding protein-1c (SREBP-1c), which is an activator of two hepatic TG synthesis enzymes, namely the diacylglycerol acetyl-transferase (DGAT) and phosphatidic acid phosphohydrolase (PA). Lowering SREBP-1c activity reduces TG production, and secretion of VLDL (Nakamura *et al.*, 2004; Harris and Bulchandani, 2006); (2) n-3 PUFAs can upregulate β -oxidation of the fatty acid substrates of hepatic TG synthesis (Pirillo and Catapano, 2015) via interaction with peroxisome proliferator-activated receptor- α (PPAR α), which is a key regulatory transcription factor for β -oxidation of fatty acids in mitochondria and peroxisome (Nakamura *et al.*, 2004), eventually, the lack of essential substrates suppresses the hepatic TG synthesis (Shearer *et al.*, 2012); and (3) n-3 PUFAs accelerate hepatic TG clearance by inducing lipolysis via insulin-promoted lipoprotein lipase (LPL) (Park and Harris, 2003). Though the pathways involved in n-3 PUFAs-mediated lowering of TG are well-known, the mechanism of n-3 PUFA's interaction with the related enzymes is not clear (Table 1).

Inflammatory response

A study showed that the levels of inflammatory cytokines, including IL-6, IL-10, and TNF- α are positively related to TG level (Gonzalez *et al.*, 2018). Since n-3 PUFAs can reduce TG level, an association between n-3 PUFAs and inflammatory response was explored. De Caterina and Libby (1996) showed that n-3 PUFAs supplementation significantly reduced the IL-1 α -induced levels of IL-6 and IL-8 in human vein endothelial cells, thus suppressing inflammatory response from IL-1 α . Another study showed that n-3 PUFA pre-treatment dramatically reduced the LPS-induced level of IL-10 in RAW 264.7 cells (Babcock *et al.*, 2002). Recently, a study suggested that n-3 PUFAs can remarkably

Table 1. The TG-lowering mechanisms of n-3 PUFAs.

Pathway	Effect
Inhibiting the enzymes of TG synthesis by downregulating the activity of SREBP-1c	Suppressing TG synthesis
Enhancing β -oxidation of fatty acids by upregulating the activity of PPAR α	Reducing the substrates for TG synthesis
Promoting the action of LPL	Increasing TG clearance

downregulate the gene expression of inflammatory cytokines IL-1 β and TNF- α in THP-1 macrophages (Allam-Ndoul *et al.*, 2016). Similarly, a rat study showed that high-fat diet-induced levels of TNF- α and IL-1 β were significantly alleviated after treatment with n-3 PUFAs (Breetha and Ramaprasad, 2018). In the apical periodontitis (AP) rat model too, n-3 PUFAs treatment led to a decrease of inflammatory cytokines TNF- α , IL-6, IL-1 β , and IL-17 (Azuma *et al.*, 2018). Moreover, a clinical trial on 324 obese subjects aged 20 to 40 indicated that n-3 PUFA supplementation for 8-week alleviated inflammatory response by downregulating CRP and IL-6 (Ramel *et al.*, 2010).

The inflammatory response involves the activation of various transcription factors, including NF- κ B, which plays an important role in many inflammatory signalling pathways. NF- κ B regulates some inflammatory cytokines (IL-1, IL-2, IL-6, IL-12, TNF- α , and *etc.*), chemokines (IL-8, MIP-1 α , MCP1, and *etc.*), adhesion molecules (ICAM, VCAM, E-selectin, and *etc.*), and inducible effector enzymes (iNOS and COX-2) (Ghosh and Karin, 2002). In the inactivated state, NF- κ B dimer remains bound to inhibitor protein I κ B in the cytoplasm; however, when cell is exposed to stress condition, the inhibitor I κ B is phosphorylated, thus releasing NF- κ B. The activated NF- κ B protein is then translocated into the nucleus to promote the expression of inflammatory proteins (Perkins, 2007).

Lo *et al.* (1999) showed that n-3 PUFA supplementation significantly inhibited the LPS-induced expression of TNF- α and NF- κ B in RAW 264.7 cells. Novak *et al.* (2003) showed that n-3 PUFAs promoted the inactivation of NF- κ B by reducing the phosphorylation of I κ B, which in turn reduced the LPS-induced level of TNF- α in RAW 264.7 cells. In LPS-exposed human monocytic THP-1 cells too, n-3 PUFAs treatment inhibited the nuclear translocation of NF- κ B by suppressing the degradation of I κ B, which in turn reduced the LPS-induced level of TNF- α (Zhao *et al.*, 2004). Similar observations were also made in animal models. Hudert *et al.* (2006) suggested that enhanced levels of n-3 PUFAs downregulated the activity of NF- κ B, thus reducing the generation of TNF- α and IL-1 β in the mice colitis model. Another study in the rat colitis model showed that n-3 PUFAs may alleviate inflammatory response via inhibition of NF- κ B which suppresses the activity of inflammatory cells (Triantafyllidis *et al.*, 2015). In the hepatic ischemia/reperfusion (I/R)-injury rat model, the hyperactivated NF- κ B and increased level of TNF- α and IL-1 β were both reversed by n-3 PUFAs via

enhanced stability of I κ B (Zuniga *et al.*, 2011). In the testicular I/R-injury rat model too, n-3 PUFAs could attenuate inflammatory response by regulating the activity of NF- κ B (Qi *et al.*, 2017). A human clinical trial, involving the patients with sickle cell disease (SCD), showed that 1-year intervention with n-3 PUFAs downregulated the level of inflammatory cytokines and NF- κ B in the patients as compared to the control group (Daak *et al.*, 2015). Overall, these findings illustrate that n-3 PUFA-mediated regulation of inflammatory response depends on the I κ B-mediated inhibition of NF- κ B.

Oxidative stress

Intracellular metabolism generates reactive oxygen species (ROS) including superoxide, hydroxyl radical, and singlet oxygen which are highly unstable and reactive molecules, and can impair the cellular components such as proteins, lipids, and nucleic acids. Therefore, intracellular antioxidant enzymes such as superoxide dismutase (SOD), glutathione peroxidase (GPx), and catalase (CAT) eliminate ROS and function as preventative measures. An imbalance of ROS production and antioxidant enzymes, named oxidative stress, can damage cellular function, thus causing cell death (Zhang *et al.*, 2016). Intracellular ROS can be upregulated by inflammatory cytokines such as TNF- α and IL-1 β (Clauzure *et al.*, 2014; Roberge *et al.*, 2014). Also, ROS can trigger the activation of NF- κ B by inducing phosphorylation and/or degradation of I κ B (Zhang *et al.*, 2016). Upregulated inflammatory cytokines promote oxidative stress, which further aggravates the inflammatory reaction via the NF- κ B pathway.

n-3 PUFA supplementation can reduce the level of ROS inducing inflammatory cytokines, thus showing a kind of antioxidant effect. Che *et al.* (2018) showed that n-3 PUFAs significantly enhanced the level of SOD to protect rat pheochromocytoma (P12) cells from oxidative damage. Under oxidative stress, cardiomyocytes (H9c2) cells showed an increased level of MDA (an end-product of oxidative damage), while the levels of antioxidant enzymes SOD, GPx, and CAT decreased. n-3 PUFA treatment also effectively ameliorated the negative effect of oxidative stress in H9c2 cells (Varghese *et al.*, 2017). In ESC-derived cardiac lineage cells, pre-treatment with n-3 PUFAs dramatically inhibited the H₂O₂-induced oxidative stress (Shabani *et al.*, 2019). The antioxidant effect of n-3 PUFAs has also been verified in a human clinical trial. Mas *et al.* (2010) showed that overweight participants who received n-3 PUFAs 4 gm/day for

six weeks exhibited a remarkable decrease in plasma F2-isoprostane (a marker of oxidative damage) level, as compared to those who received olive oil. A randomised controlled trial of 105 overweight subjects showed that as compared to the control group, the n-3 PUFA treatment group had a lower level of F2-isoprostane during the 4-month follow-up period (Kiecolt-Glaser *et al.*, 2013). Recently, another study in type 2 diabetic patients suggested that MDA and F2-isoprostane levels were remarkably reduced after administration with n-3 PUFAs for two weeks (Vericel *et al.*, 2015).

Although the above studies demonstrated that n-3 PUFAs can attenuate oxidative stress-induced damage, the mechanism remains unclear. Since mitochondria and NADPH oxidases are the main sources of ROS, mitochondrial dysfunction is considered the main causative factor of oxidative stress. Emerging evidence indicates that inflammatory response is a result of mitochondrial abnormality which aggravates intracellular ROS levels (Dan Dunn *et al.*, 2015; Angelova and Abramov, 2018). A study showed that n-3 PUFAs can effectively ameliorate oxidative damage and mitochondrial dysfunction both *in vivo* and *in vitro* (Zhang *et al.*, 2018a). n-3 PUFAs could improve oxidative damage by inhibiting the activity of mitochondrial respiratory chain enzymes in rats' brain tissue under oxidative stress (Carvalho-Silva *et al.*, 2019). Nuclear factor E2-related factor 2 (Nrf2) is a transcription factor that regulates the expression of various antioxidant enzymes including haem oxygenase 1 (HO-1). Under normal conditions, Nrf2 is silenced by Keap1 (Kelch-like ECH-associated protein 1), but under oxidative stress, it gets released

from Keap1 for nuclear translocation, where it upregulates the transcription of antioxidant enzymes (Tonelli *et al.*, 2018). Zhang *et al.* (2014) suggested that protective effects of n-3 PUFAs were in part due to the activation of Nrf2 in the brain injury mouse model. In another study, hepatic injury or a high-fat diet significantly downregulated Nrf2/HO-1 in mice, whereas n-3 PUFA treatment dramatically alleviated hepatic injury by enhancing the Nrf2/HO-1 activity (Gonzalez *et al.*, 2018). Similarly, Yang *et al.* (2013) showed that n-3 PUFAs enhanced nuclear translocation of Nrf2, thus promoting the latter's activity. All these findings suggest that n-3 PUFAs show antioxidant effect via two major pathways: (1) by reducing ROS production from mitochondrial dysfunction, and (2) by eliminating the redundant ROS by Nrf2 activation (Figure 1).

Protective effect of n-3 PUFAs

Several common diseases of the modern world have emerged as serious health issues. In 2016, about 121.5 million young adults (≥ 20 years) had more than one type of cardiovascular disease (CVD), and approximately 17.6 million deaths were attributed to CVD worldwide. DM is another prevalent disease. From 2013 to 2016, about 9.8% of US adults (≈ 26 million) were diagnosed with DM, costing ~ 327 billion dollars to patients in 2017 (Benjamin *et al.*, 2019). Similarly, non-alcoholic fatty liver disease (NAFLD) is also on a rising trajectory affecting around 25.24% of the global population in 2016 (Younossi *et al.*, 2016). High TG level, inflammation, and oxidative stress are general clinical characteristics of these disorders. High TG level is positively linked to endothelial dysfunction,

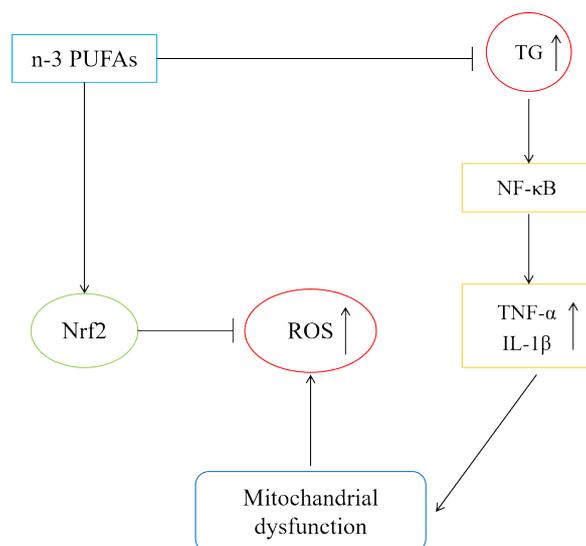


Figure 1. An unhealthy diet and lifestyle-induced high TG level can promote a series of intracellular negative reactions, while n-3 PUFA supplementation effectively reduces the enhanced TG level, and alleviates oxidative damage by promoting Nrf2 activation.

an important contributor to cardiovascular events, which is further aggravated by inflammation and oxidative stress (Steven *et al.*, 2019). Similarly, high TG level can trigger DM by impairing insulin synthesis due to TG deposition-induced abnormality of insulin β -cells (Raz *et al.*, 2005). Kozakova *et al.* (2019) found that as compared to healthy individuals, type 2 diabetes (T2D) patients exhibited higher plasma levels of IL-6 and IL-8. Notably, NAFLD is also regarded as a consequence of excessive TG deposition in the liver, which can further deteriorate to non-alcoholic steatohepatitis (NASH). After a series of inflammatory events and oxidative damages, NASH can finally develop into hepatic fibrosis (Peverill *et al.*, 2014). Overall, these findings highlight that common lifestyle diseases are aggravated by TG levels, which can be countered with n-3 PUFA supplementation.

Cardiovascular disease (CVD)

In the 1970s, some Danish physicians found that the risk of coronary heart disease (CHD) among Greenland Eskimos was significantly lower than those who lived in Denmark. They observed that n-3 PUFA-rich fish diet led to a higher concentration of n-3 PUFAs in Eskimos blood (Bang *et al.*, 1976). Subsequently, more epidemiological studies were performed to validate this claim. In 1985, Kromhout *et al.* (1985) showed that CHD-related mortality was about 50% lower in those who eat fish at least 30 g/day than those who rarely consumed fish. This was also supported by the American nurses' health study which started in 1976 with a 16-year follow-up and included 84,688 healthy women nurses, aged 30 to 55. The study showed an inverse association between CHD risk and fish consumption (Hu *et al.*, 2002). Another 3-year follow-up study among 18,244 healthy men (aged 45 to 64) in Shanghai, China suggested a significant link between fish diet and lower risk of fatal myocardial infarction (MI) (Yuan *et al.*, 2001). The same was reported in Japan Public Health Centre-based (JPHC) study, which supervised 41,578 Japanese middle-aged individuals between 1990 and 2001. The study found a lower CVD risk among more fish-consuming individuals than those who had less fish (Iso *et al.*, 2006).

Apart from the above-discussed studies, several clinical trials also established the protective effect of n-3 PUFAs. In 2018, an RCT (randomised controlled trial) of 421,309 participants who were free of CVD risk showed a 10% decline of CVD mortality in those who consumed more fish than those who consumed less (Zhang *et al.*, 2018b). Another RCT of 427,678 healthy UK individuals in

2020 indicated a dramatic correlation between higher n-3 PUFAs intake and lower CVD events, including CVD-related mortality (Li *et al.*, 2020).

Additionally, the beneficial effect of n-3 PUFAs as secondary prevention measure was also investigated. A GISSI-Prevenzione trial of 11,323 MI survivors for a 3.5-year follow-up study showed that n-3 PUFA treatment reduced CVD mortality by about 30% as compared to the placebo (Marchioli *et al.*, 2002). A JELIS trial with a 5-year follow-up of patients with a history of CVD showed that statin supplemented with EPA reduced CVD events by 19% than statin alone (Yokoyama *et al.*, 2007). Most recently in 2019, a 4.9-year follow-up RCT of 8,179 hypertriglyceridemia patients suggested that CVD events among patients treated with EPA supplementation were remarkably lower (71% in secondary prevention trial) than those in the placebo group (Bhatt *et al.*, 2019).

Meta-analyses of RCTs showed a significant preventive effect of n-3 PUFAs. A meta-analysis of 11 RCTs, including 39,044 patients with a history of CVD, found that the patients who took 1.8 gm/day EPA/DHA exhibited a significantly lower CVD risk than those in the control group (Marik and Varon, 2009). In 2013, a meta-analysis of 11 RCTs investigated the effect of n-3 PUFA supplementation (1 gm/day for at least one year), and found that as compared to those who took a placebo, 32% reduction of cardiac death, 33% reduction of sudden death, and 25% reduction of MI was noticed in those who received n-3 PUFAs (Casula *et al.*, 2013). Another meta-analysis of 14 RCTs in 2014, including 32,656 individuals with CHD, showed a 7% reduction in CVD events, 12% reduction in death from cardiac causes, 14% reduction in sudden cardiac death, and 8% reduction in all-cause mortality among patients who received n-3 PUFAs as compared to the control group (Wen *et al.*, 2014). Recently, a 2019 meta-analysis of 13 RCTs, containing 127,477 participants, evaluated dose-dependent benefits of n-3 PUFAs in CVD events, and found that n-3 PUFA supplementation significantly lowered the CVD risk, and showed an inverse linear dose-response relationship in the range of 0 - 4,000 mg/d of n-3 PUFAs (Hu *et al.*, 2019).

Diabetes mellitus (DM)

So far, various clinical trials have shown the beneficial effect of n-3 PUFAs in DM. Wang *et al.* (2003) measured the plasma fatty acid composition from 2,909 participants (aged 45 to 64) in a 9-year follow-up study, and found that n-3 PUFAs levels were significantly lower in diabetics than in healthy

participants, suggesting a possible association between low n-3 PUFA levels and increased risk of DM. In 2004, a study examined the relationship between diet and type 2 diabetes (T2D) prevalence in the Nordic countries, and showed that lower T2D incidence was linked to higher dietary n-3 PUFA content (Thorsdottir *et al.*, 2004). In 2011, an 8 to 9-year follow-up health study of 64,193 Shanghai women, who were free of T2D, CVD, and cancer, reported that n-3 PUFA intake was inversely associated with T2D risk (Villegas *et al.*, 2011). Similarly, a meta-analysis of 24 RCTs suggested a significant inverse correlation between n-3 PUFA intake and T2D risk in Asians, while n-3 PUFA content was dramatically lower in T2D patients than in healthy population (Zheng *et al.*, 2012).

A 2013 study, which was designed to examine the effect of n-3 PUFAs on glucose metabolism in elderly T2D patients for 3-month, showed that those with n-3 PUFA supplementation had a lower fasting plasma glucose (FPG) level, haemoglobin A1c (HbA1c), remnant like particle (RLP), and apolipoprotein B (apo B) as compared to the control individuals, thus suggesting a significant improvement in impaired-glucose metabolism in elderly T2D patients (Ogawa *et al.*, 2013). Similarly, Kurt *et al.* (2016) showed a significant decline in FPG, HbA1c, and pentosidine among T2D patients who received n-3 PUFAs (1.2 gm/day) for 2-month. A 2018 meta-analysis of 5 RCTs suggested that n-3 PUFA supplementation effectively reduced the level of FPG, insulin resistance (IR), and C-reactive protein (CRP) among patients with gestational diabetes, which occurs during pregnancy (Zhong and Wang, 2019).

Non-alcoholic fatty liver disease (NAFLD)

The beneficial effect of n-3 PUFAs in NAFLD has been supported by many clinical trials. In 2015, an RCT of 51 paediatric patients with NAFLD showed that n-3 PUFA supplementation for six months inhibited lipid accumulation as compared to the placebo group (Pacifico *et al.*, 2015). Li *et al.* (2015) showed a significant reduction of plasma alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels in NASH patients who received n-3 PUFAs for six months, along with a significant reduction in TAG, CRP, and MDA levels. This study clearly showed that n-3 PUFAs can ameliorate impaired liver, and inhibit inflammation in NASH patients.

Recently, a meta-analysis of seven RCTs involving 442 patients on n-3 PUFAs effect on

NAFLD showed that TAG, TC, and LDL-C levels were significantly reduced in the n-3 PUFAs group as compared to the control group; meanwhile, significant reduction of ALT, AST, and GGT was also noticed as a secondary effect which attenuated fatty liver and fibrosis (He *et al.*, 2016). Similarly, a meta-analysis of four RCT involving 263 children with NAFLD demonstrated that n-3 PUFAs inhibited the progression of hepatic steatosis (Chen *et al.*, 2018). Another meta-analysis of eight RCTs involving 1424 participants with NAFLD showed that n-3 PUFA supplementation reduced liver fat, and improved liver function (Yan *et al.*, 2018). A 2018 meta-analysis of 11 RCTs showed that 1 gm/day supplementation of n-3 PUFAs in NAFLD patients resulted in 3.14 U/L, 2.43 U/L, 2.74%, and 9.97 mg/dL decline in the levels of ALT, AST, liver fat, and TAG, respectively (Guo *et al.*, 2018). Although many studies showed the protective ability of n-3 PUFAs in NAFLD patients, larger scale and longer follow-up studies are needed to further validate these results.

Conclusion

Hypertriglyceridemia has become a prevalent disease due to high-fat diet and less exercise in the modern society. High TG level is a serious risk factor that enhances the incidence of CVD, DM, and NAFLD. In addition, high TG levels can promote inflammation and oxidative damage, which can aggravate the progression of the diseases. Several recent studies, including human clinical trials, suggest that n-3 PUFA supplementation can ameliorate these risks. It is speculated that n-3 PUFAs alleviate inflammation and oxidative stress by downregulating the activity of NF- κ B and Nrf2. Besides, n-3 PUFAs reduce TG level by inhibiting synthesis or enhancing the clearance of TG. Based on these findings, one can suggest that n-3 PUFA supplements or n-3 PUFA-rich marine-derived foods can have beneficial health effects, and therefore should be included in the daily diet.

Acknowledgement

The present work was financially supported by the National Science Fund for Distinguished Young Scholars of China (grant no.: 31725022), and the Science and Technology Planning Project of Guangdong Province (grant no.: 2019A050503002). The authors are thankful to Guangdong Yue-s Special Nutrition Technology Co., Ltd. and the School of Food Sciences and Engineering, South China University of Technology.

References

- Allam-Ndoul, B., Guenard, F., Barbier, O. and Vohl, M. C. 2016. Effect of n-3 fatty acids on the expression of inflammatory genes in THP-1 macrophages. *Lipids in Health and Disease* 15: article no. 69.
- Alves-Bezerra, M. and Cohen, D. E. 2017. Triglyceride metabolism in the liver. *Comprehensive Physiology* 8: 1-8.
- Angelova, P. R. and Abramov, A. Y. 2018. Role of mitochondrial ROS in the brain: from physiology to neurodegeneration. *FEBS Letters* 592: 692-702.
- Azuma, M. M., Gomes-Filho, J. E., Ervolino, E., Cardoso, C. B. M., Pipa, C. B., Kawai, T., ... and Cintra, L. T. A. 2018. Omega-3 fatty acids reduce inflammation in rat apical periodontitis. *Journal of Endodontics* 44: 604-608.
- Babcock, T. A., Novak, T., Ong, E., Jho, D. H., Helton, W. S. and Espot, N. J. 2002. Modulation of lipopolysaccharide-stimulated macrophage tumor necrosis factor- α production by ω -3 fatty acid is associated with differential cyclooxygenase-2 protein expression and is independent of interleukin-10. *Journal of Surgical Research* 107: 135-139.
- Bang, H. O., Dyerberg, J. and Hjoorne, N. 1976. The composition of food consumed by Greenland Eskimos. *Acta Medica Scandinavica* 200: 69-73.
- Benjamin, E. J., Muntner, P. and Alonso, A. 2019. Heart disease and stroke statistics - 2019 update: a report from the American Heart Association. *Circulation* 139: e56-e528.
- Bhatt, D. L., Steg, P. G., Miller, M., Brinton, E. A., Jacobson, T. A., Ketchum, S. B., ... and Ballantyne, C. M. 2019. Cardiovascular risk reduction with icosapent ethyl for hypertriglyceridemia. *The New England Journal of Medicine* 380: 11-22.
- Breetha, R. and Ramaprasad, T. 2018. Dietary n-3 but not n-6 fatty acids down-regulate maternal dyslipidemia induced inflammation: a three-generation study in rats. *Prostaglandins, Leukotrienes and Essential Fatty Acids* 135: 83-91.
- Calder, P. C. 2012. Mechanisms of action of (n-3) fatty acids. *Journal of Nutrition* 142: 592S-599S.
- Carvalho-Silva, M., Gomes, L. M., Gomes, M. L., Ferreira, B. K., Schuck, P. F., Ferreira, G. C., ... and Streck, E. L. 2019. Omega-3 fatty acid supplementation can prevent changes in mitochondrial energy metabolism and oxidative stress caused by chronic administration of L-tyrosine in the brain of rats. *Metabolic Brain Disease* 34: 1207-1219.
- Casula, M., Soranna, D., Catapano, A. L. and Corrao, G. 2013. Long-term effect of high dose omega-3 fatty acid supplementation for secondary prevention of cardiovascular outcomes: a meta-analysis of randomized, placebo controlled trials. *Atherosclerosis* 14(2): 243-251.
- Chan, D. C., Pang, J., Barrett, P. H., Sullivan, D. R., Burnett, J. R., van Bockxmeer, F. M. and Watts, G. F. 2016. Omega-3 fatty acid ethyl esters diminish postprandial lipemia in familial hypercholesterolemia. *The Journal of Clinical Endocrinology and Metabolism* 101: 3732-3739.
- Che, H., Fu, X., Zhang, L., Gao, X., Wen, M., Du, L., ... and Wang, Y. 2018. Neuroprotective effects of n-3 polyunsaturated fatty acid-enriched phosphatidylserine against oxidative damage in PC12 cells. *Cellular and Molecular Neurobiology* 38: 657-668.
- Chen, L. H., Wang, Y. F., Xu, Q. H. and Chen, S. S. 2018. Omega-3 fatty acids as a treatment for non-alcoholic fatty liver disease in children: a systematic review and meta-analysis of randomized controlled trials. *Clinical Nutrition* 37: 516-521.
- Clazure, M., Valdivieso, A. G., Massip Copiz, M. M., Schulman, G., Teiber, M. L. and Santa-Coloma, T. A. 2014. Disruption of interleukin-1 β autocrine signaling rescues complex I activity and improves ROS levels in immortalized epithelial cells with impaired cystic fibrosis transmembrane conductance regulator (CFTR) function. *PLoS One* 9: article ID e99257.
- Daak, A. A., Elderderly, A. Y., Elbashir, L. M., Mariniello, K., Mills, J., Scarlett, G., ... and Ghebremeskel, K. 2015. Omega 3 (n-3) fatty acids down-regulate nuclear factor- κ B (NF- κ B) gene and blood cell adhesion molecule expression in patients with homozygous sickle cell disease. *Blood Cells, Molecules and Diseases* 55: 48-55.
- Dan Dunn, J., Alvarez, L. A., Zhang, X. and Soldati, T. 2015. Reactive oxygen species and mitochondria: a nexus of cellular homeostasis. *Redox Biology* 6: 472-485.
- De Caterina, R. and Libby, P. 1996. Control of endothelial leukocyte adhesion molecules by fatty acids. *Lipids* 31: S57-S63.
- Ghosh, S. and Karin, M. 2002. Missing pieces in the NF- κ B puzzle. *Cell* 109(2): S81-S96.
- Gonzalez, M. B., Lane, M., Knight, E. J. and Robker, R. L. 2018. Inflammatory markers in human follicular fluid correlate with lipid levels and

- body mass index. *Journal of Reproductive Immunology* 130: 25-29.
- Guo, X. F., Yang, B., Tang, J. and Li, D. 2018. Fatty acid and non-alcoholic fatty liver disease: meta-analyses of case-control and randomized controlled trials. *Clinical Nutrition* 37: 113-122.
- Harris, W. S. and Bulchandani, D. 2006. Why do omega-3 fatty acids lower serum triglycerides? *Current Opinion in Lipidology* 17: 387-393.
- He, X. X., Wu, X. L., Chen, R. P., Chen, C., Liu, X. G., Wu, B. J. and Huang, Z. M. 2016. Effectiveness of omega-3 polyunsaturated fatty acids in non-alcoholic fatty liver disease: a meta-analysis of randomized controlled trials. *PLoS One* 11: article ID e0162368.
- Hu, F. B., Bronner, L., Willett, W. C., Stampfer, M. J., Rexrode, K. M., Albert, C. M., ... and Manson, J. E. 2002. Fish and omega-3 fatty acid intake and risk of coronary heart disease in women. *JAMA* 287: 1815-1821.
- Hu, Y., Hu, F. B. and Manson, J. E. 2019. Marine omega-3 supplementation and cardiovascular disease: an updated meta-analysis of 13 randomized controlled trials involving 127,477 participants. *Journal of the American Heart Association* 8: article ID e013543.
- Hudert, C. A., Weylandt, K. H., Lu, Y., Wang, J., Hong, S., Dignass, A., ... and Kang, J. X. 2006. Transgenic mice rich in endogenous omega-3 fatty acids are protected from colitis. *Proceedings of the National Academy of Sciences of the United States of America* 103: 11276-11281.
- Innes, J. K. and Calder, P. C. 2020. Marine omega-3 (n-3) fatty acids for cardiovascular health: an update for 2020. *International Journal of Molecular Sciences* 21(4): article no. 1362.
- Iso, H., Kobayashi, M., Ishihara, J., Sasaki, S., Okada, K., Kita, Y., ... and Tsugane, S. 2006. Intake of fish and n3 fatty acids and risk of coronary heart disease among Japanese. *Circulation* 113: 195-202.
- Kiecolt-Glaser, J. K., Epel, E. S., Belury, M. A., Andridge, R., Lin, J., Glaser, R., ... and Blackburn, E. 2013. Omega-3 fatty acids, oxidative stress, and leukocyte telomere length: a randomized controlled trial. *Brain, Behavior, and Immunity* 28: 16-24.
- Kozakova, M., Morizzo, C., Goncalves, I., Natali, A., Nilsson, J. and Palombo, C. 2019. Cardiovascular organ damage in type 2 diabetes mellitus: the role of lipids and inflammation. *Cardiovascular Diabetology* 18: article no. 61.
- Kris-Etherton, P. M., Innis, S., American Dietetic Association and Dietitians of Canada. 2007. Position of the American Dietetic Association and Dietitians of Canada: dietary fatty acids. *Journal of the American Dietetic Association* 107(9): 1599-1611.
- Kromhout, D., Bosschieter, E. B. and de Lezenne Coulander, C. 1985. The inverse relation between fish consumption and 20-year mortality from coronary heart disease. *The New England Journal of Medicine* 312: 1205-1209.
- Kurt, A., Andican, G., Siva, Z. O., Andican, A. and Burcak, G. 2016. The effects of n-3 long-chain polyunsaturated fatty acid supplementation on AGEs and sRAGE in type 2 diabetes mellitus. *Journal of Physiology and Biochemistry* 72: 679-687.
- Li, Y. H., Yang, L. H., Sha, K. H., Liu, T. G., Zhang, L. G. and Liu, X. X. 2015. Efficacy of poly-unsaturated fatty acid therapy on patients with nonalcoholic steatohepatitis. *World Journal of Gastroenterology* 21: 7008-7013.
- Li, Z. H., Zhong, W. F., Liu, S., Kraus, V. B., Zhang, Y. J., Gao, X., ... and Mao, C. 2020. Associations of habitual fish oil supplementation with cardiovascular outcomes and all cause mortality: evidence from a large population based cohort study. *BMJ* 368: article no. m456.
- Lo, C. J., Chiu, K. C., Fu, M., Lo, R. and Helton, S. 1999. Fish oil decreases macrophage tumor necrosis factor gene transcription by altering the NFκB activity. *Journal of Surgical Research* 82: 216-221.
- Marchioli, R., Barzi, F., Bomba, E., Chieffo, C., Di Gregorio, D., Di Mascio, R., ... and Valagussa, F. 2002. Early protection against sudden death by n-3 polyunsaturated fatty acids after myocardial infarction: time-course analysis of the results of the Gruppo Italiano per lo Studio della Sopravvivenza nell'Infarto Miocardico (GISSI) - Prevenzione. *Circulation* 105: 1897-1903.
- Marik, P. E. and Varon, J. 2009. Omega-3 dietary supplements and the risk of cardiovascular events: a systematic review. *Clinical Cardiology* 32: 365-372.
- Mas, E., Woodman, R. J., Burke, V., Puddey, I. B., Beilin, L. J., Durand, T. and Mori, T. A. 2010. The omega-3 fatty acids EPA and DHA decrease plasma F(2)-isoprostanes: results from two placebo-controlled interventions. *Free Radical Research* 44: 983-990.
- Mozaffarian, D. and Wu, J. H. 2011. Omega-3 fatty acids and cardiovascular disease: effects on risk factors, molecular pathways, and clinical events. *Journal of the American College of Cardiology*

- 58: 2047-2067.
- Nakamura, M. T., Cheon, Y., Li, Y. and Nara, T. Y. 2004. Mechanisms of regulation of gene expression by fatty acids. *Lipids* 39: 1077-1083.
- Nordestgaard B. G. 2016. Triglyceride-rich lipoproteins and atherosclerotic cardiovascular disease: new insights from epidemiology, genetics, and biology. *Circulation Research* 118(4): 547-563.
- Novak, T. E., Babcock, T. A., Jho, D. H., Helton, W. S. and Espot, N. J. 2003. NF- κ B inhibition by ω -3 fatty acids modulates LPS-stimulated macrophage TNF- α transcription. *American Journal of Physiology - Lung Cellular and Molecular Physiology* 284(1): L84-L89.
- Ogawa, S., Abe, T., Nako, K., Okamura, M., Senda, M., Sakamoto, T. and Ito, S. 2013. Eicosapentaenoic acid improves glycemic control in elderly bedridden patients with type 2 diabetes. *The Tohoku Journal of Experimental Medicine* 231: 63-74.
- Oh, P. C., Koh, K. K., Sakuma, I., Lim, S., Lee, Y., Lee, S., ... and Shin, E. K. 2014. Omega-3 fatty acid therapy dose-dependently and significantly decreased triglycerides and improved flow-mediated dilation, however, did not significantly improve insulin sensitivity in patients with hypertriglyceridemia. *International Journal of Cardiology* 176: 696-702.
- Pacifico, L., Bonci, E., Di Martino, M., Versacci, P., Andreoli, G., Silvestri, L. M. and Chiesa, C. 2015. A double-blind, placebo-controlled randomized trial to evaluate the efficacy of docosahexaenoic acid supplementation on hepatic fat and associated cardiovascular risk factors in overweight children with nonalcoholic fatty liver disease. *Nutrition, Metabolism and Cardiovascular Diseases* 25: 734-741.
- Park, Y. and Harris, W. S. 2003. Omega-3 fatty acid supplementation accelerates chylomicron triglyceride clearance. *Journal of Lipid Research* 44: 455-463.
- Perkins, N. D. 2007. Integrating cell-signalling pathways with NF- κ B and IKK function. *Nature Reviews Molecular Cell Biology* 8: 49-62.
- Peverill, W., Powell, L. W. and Skoien, R. 2014. Evolving concepts in the pathogenesis of NASH: beyond steatosis and inflammation. *International Journal of Molecular Sciences* 15: 8591-8638.
- Pirillo, A. and Catapano, A. L. 2015. Update on the management of severe hypertriglyceridemia--focus on free fatty acid forms of omega-3. *Drug Design, Development and Therapy* 9: 2129-2137.
- Qi, X., Qin, Z., Tang, J., Han, P., Xing, Q., Wang, K., ... and Zhang, W. 2017. Omega-3 polyunsaturated fatty acids ameliorates testicular ischemia-reperfusion injury through the induction of Nrf2 and inhibition of NF- κ B in rats. *Experimental and Molecular Pathology* 103(1): 44-50.
- Ramel, A., Martinez, J. A., Kiely, M., Bandarra, N. M., and Thorsdottir, I. 2010. Effects of weight loss and seafood consumption on inflammation parameters in young, overweight and obese European men and women during 8 weeks of energy restriction. *European Journal of Clinical Nutrition* 64: 987-993.
- Raz, I., Eldor, R., Cernea, S. and Shafrir, E. 2005. Diabetes: insulin resistance and derangements in lipid metabolism. Cure through intervention in fat transport and storage. *Diabetes/Metabolism Research and Reviews* 21: 3-14.
- Reiner, Z. 2017. Hypertriglyceridaemia and risk of coronary artery disease. *Nature Reviews Cardiology* 14: 401-411.
- Rhoads, J. P. and Major, A. S. 2018. How oxidized low-density lipoprotein activates inflammatory responses. *Critical Reviews in Immunology* 38: 333-342.
- Roberge, S., Roussel, J., Andersson, D. C., Meli, A. C., Vidal, B., Blandel, F., ... and Fauconnier, J. 2014. TNF- α -mediated caspase-8 activation induces ROS production and TRPM2 activation in adult ventricular myocytes. *Cardiovascular research* 103: 90-99.
- Shabani, P., Ghazizadeh, Z., Gorgani-Firuzjaee, S., Molazem, M., Rajabi, S., Vahdat, S., ... and Baharvand, H. 2019. Cardioprotective effects of omega-3 fatty acids and ascorbic acid improve regenerative capacity of embryonic stem cell-derived cardiac lineage cells. *BioFactors* 45: 427-438.
- Shahidi, F. and Ambigaipalan, P. 2018. Omega-3 polyunsaturated fatty acids and their health benefits. *Annual Review of Food Science and Technology* 9: 345-381.
- Shearer, G. C., Savinova, O. V. and Harris, W. S. 2012. Fish oil -- how does it reduce plasma triglycerides? *Biochimica et Biophysica Acta* 1821: 843-851.
- Steven, S., Frenis, K., Oelze, M., Kalinovic, S., Kuntic, M., Bayo Jimenez, M. T., ... and Daiber, A. 2019. Vascular inflammation and oxidative stress: major triggers for cardiovascular disease. *Oxidative Medicine and Cellular Longevity* 2019: article ID 7092151.
- Thorsdottir, I., Hill, J. and Ramel, A. 2004. Omega-3 fatty acid supply from milk associates with lower

- type 2 diabetes in men and coronary heart disease in women. *Preventive Medicine* 39: 630-634.
- Tonelli, C., Chio, I. I. C. and Tuveson, D. A. 2018. Transcriptional regulation by Nrf2. *Antioxidants and Redox Signaling* 29: 1727-1745.
- Toth, P. P. 2016. Triglyceride-rich lipoproteins as a causal factor for cardiovascular disease. *Vascular Health and Risk Management* 12: 171-183.
- Triantafyllidis, I., Poutahidis, T., Taitzoglou, I., Kesisoglou, I., Lazaridis, C. and Botsios, D. 2015. Treatment with Mesna and n-3 polyunsaturated fatty acids ameliorates experimental ulcerative colitis in rats. *International Journal of Experimental Pathology* 96: 433-443.
- Varghese, M. V., Abhilash, M., Paul, M. V., Alex, M. and Nair, R. H. 2017. Omega-3 fatty acid protects against arsenic trioxide-induced cardiotoxicity *in vitro* and *in vivo*. *Cardiovascular Toxicology* 17: 109-119.
- Vericel, E., Colas, R., Calzada, C., Le, Q. H., Feugier, N., Cugnet, C., ... and Lagarde, M. 2015. Moderate oral supplementation with docosahexaenoic acid improves platelet function and oxidative stress in type 2 diabetic patients. *Thrombosis and Haemostasis* 114: 289-296.
- Villegas, R., Xiang, Y. B., Elasy, T., Li, H. L., Yang, G., Cai, H., ... and Shu, X.-O. 2011. Fish, shellfish, and long-chain n-3 fatty acid consumption and risk of incident type 2 diabetes in middle-aged Chinese men and women. *The American Journal of Clinical Nutrition* 94: 543-551.
- Wang, L., Folsom, A. R., Zheng, Z. J., Pankow, J. S. and Eckfeldt, J. H. 2003. Plasma fatty acid composition and incidence of diabetes in middle-aged adults: the Atherosclerosis Risk in Communities (ARIC) study. *The American Journal of Clinical Nutrition* 78: 91-98.
- Wen, Y. T., Dai, J. H. and Gao, Q. 2014. Effects of omega-3 fatty acid on major cardiovascular events and mortality in patients with coronary heart disease: a meta-analysis of randomized controlled trials. *Nutrition, Metabolism and Cardiovascular Diseases* 24: 470-475.
- Yan, J. H., Guan, B. J., Gao, H. Y. and Peng, X. E. 2018. Omega-3 polyunsaturated fatty acid supplementation and non-alcoholic fatty liver disease: a meta-analysis of randomized controlled trials. *Medicine* 97: article ID e12271.
- Yang, Y. C., Lii, C. K., Wei, Y. L., Li, C. C., Lu, C. Y., Liu, K. L. and Chen, H. W. 2013. Docosahexaenoic acid inhibition of inflammation is partially via cross-talk between Nrf2/heme oxygenase 1 and IKK/NF- κ B pathways. *Journal of Nutritional Biochemistry* 24: 204-212.
- Yokoyama, M., Origasa, H., Matsuzaki, M., Matsuzawa, Y., Saito, Y., Ishikawa, Y., ... and Shirato, K. 2007. Effects of eicosapentaenoic acid on major coronary events in hypercholesterolaemic patients (JELIS): a randomised open-label, blinded endpoint analysis. *Lancet* 369: 1090-1098.
- Younossi, Z. M., Koenig, A. B., Abdelatif, D., Fazel, Y., Henry, L. and Wymer, M. 2016. Global epidemiology of nonalcoholic fatty liver disease - meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* 64: 73-84.
- Yuan, J. M., Ross, R. K., Gao, Y. T. and Yu, M. C. 2001. Fish and shellfish consumption in relation to death from myocardial infarction among men in Shanghai, China. *American Journal of Epidemiology* 154: 809-816.
- Zeman, M., Zak, A., Vecka, M., Tvrzicka, E., Pisarikova, A. and Stankova, B. 2006. N-3 fatty acid supplementation decreases plasma homocysteine in diabetic dyslipidemia treated with statin-fibrate combination. *Journal of Nutritional Biochemistry* 17: 379-384.
- Zhang, J., Wang, X., Vikash, V., Ye, Q., Wu, D., Liu, Y. and Dong, W. 2016. ROS and ROS-mediated cellular signaling. *Oxidative Medicine and Cellular Longevity* 2016: article ID 4350965.
- Zhang, M., Wang, S., Mao, L., Leak, R. K., Shi, Y., Zhang, W., ... and Zhang, F. 2014. Omega-3 fatty acids protect the brain against ischemic injury by activating Nrf2 and upregulating heme oxygenase 1. *The Journal of Neuroscience* 34: 1903-1915.
- Zhang, T., Wu, P., Zhang, J. H., Li, Y., Xu, S., Wang, C., ... and Shi, H. 2018a. Docosahexaenoic acid alleviates oxidative stress-based apoptosis via improving mitochondrial dynamics in early brain injury after subarachnoid hemorrhage. *Cellular and Molecular Neurobiology* 38: 1413-1423.
- Zhang, Y., Zhuang, P., He, W., Chen, J. N., Wang, W. Q., Freedman, N. D., ... and Jiao, J. J. 2018b. Association of fish and long-chain omega-3 fatty acids intakes with total and cause-specific mortality: prospective analysis of 421,309 individuals. *Journal of Internal Medicine* 284: 399-417.
- Zhao, Y., Joshi-Barve, S., Barve, S. and Chen, L. H. 2004. Eicosapentaenoic acid prevents LPS-induced TNF- α expression by preventing NF- κ B activation. *Journal of the American College of Nutrition* 23: 71-78.
- Zheng, J. S., Huang, T., Yang, J., Fu, Y. Q. and Li, D. 2012. Marine n-3 polyunsaturated fatty acids are

inversely associated with risk of type 2 diabetes in Asians: a systematic review and meta-analysis. *PLoS One* 7: article ID e44525.

Zhong, N. and Wang, J. 2019. The efficacy of omega-3 fatty acid for gestational diabetes: a meta-analysis of randomized controlled trials. *Gynecological Endocrinology* 35: 4-9.

Zhu, F. S., Liu, S., Chen, X. M., Huang, Z. G. and Zhang, D. W. 2008. Effects of n-3 polyunsaturated fatty acids from seal oils on nonalcoholic fatty liver disease associated with hyperlipidemia. *World Journal of Gastroenterology* 14: 6395-6400.

Zuniga, J., Cancino, M., Medina, F., Varela, P., Vargas, R., Tapia, G., ... and Fernandez, V. 2011. N-3 PUFA supplementation triggers PPAR- α activation and PPAR- α /NF- κ B interaction: anti-inflammatory implications in liver ischemia-reperfusion injury. *PLoS One* 6: article ID e28502.